

is not known. One explanation could be that heparin inhibits the toxicity of stimulated PMN by its binding to receptors present on hepatocyte plasma membrane [8]. This explanation seems unlikely since we observed that, when hepatocytes were preincubated with heparin, and then washed before the addition of PMN, they were not protected from PMN toxicity ($45.2 \pm 2.9\%$ ALT release after preincubation without heparin vs $42.0 \pm 7.1\%$ after preincubation with $50 \mu\text{g/ml}$ heparin). The likely explanation is that the protective effect of heparin is due to its binding to proteinases released by stimulated PMN. Actually, it is well established that heparin is able to bind to and to inactivate proteinases released from human PMN, e.g. chymotrypsin- and elastase-like enzymes [1, 3].

PMN are known to play a major role in acute inflammatory reaction [13], and it has been suggested that heparin, besides its anticoagulant effect, had anti-inflammatory properties [12]. It is tempting to speculate that this effect of heparin might be in part explained by the inactivation of mediators released by PMN.

In summary, the effect of heparin was studied on the proteinase-mediated toxicity of human PMN against isolated rat hepatocytes. Opsonized zymosan-stimulated PMN were markedly toxic to hepatocytes and this cytotoxicity was inhibited by 32 to 55% by concentrations of heparin ranging from $2 \mu\text{g/ml}$ to $50 \mu\text{g/ml}$. This effect was not due to an inhibition of the stimulation of PMN since heparin did not decrease the release of two neutrophil lysosomal enzymes, *N*-acetyl- β -glucosaminidase and β -glucuronidase. It was explained by the inhibition of the post-secretory step of PMN toxicity since heparin reduced by 60% the toxicity of a proteinase-containing PMN lysate and by 69% that of a preparation of PMN granule proteins.

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REFERENCES

1. Marossy K, Interaction of the chymotrypsin- and elastase-like enzymes of the human granulocyte with glycosaminoglycans. *Biochim Biophys Acta* **659**: 351–361, 1981.
2. Avila JL and Convit J, Physicochemical characteristics of the glycosaminoglycan-lysosomal enzyme interaction *in vitro*. *Biochem J* **160**: 129–136, 1976.
3. Redini F, Tixier J-M, Petitou M, Choay J, Robert L and Hornebeck W, Inhibition of leucocyte elastase by heparin and its derivatives. *Biochem J* **252**: 515–519, 1988.
4. Mavier P, Préaux A-M, Guigui B, Lescs M-C, Zafrani ES and Dhumeaux D, *In vitro* toxicity of polymorphonuclear neutrophils to rat hepatocytes: Evidence for a proteinase-mediated mechanism. *Hepatology* **8**: 254–258, 1988.
5. Laperche Y, Préaux A-M, Feldmann G, Mahu J-L and Berthelot P, Effect of fasting on organic anion uptake by isolated rat liver cells. *Hepatology* **1**: 617–621, 1981.
6. Dallegri F, Frumento G and Patrone F, Mechanisms of tumour cell destruction by PMA-activated human neutrophils. *Immunology* **48**: 273–279, 1983.
7. Harlan JM, Killen PD, Harker LA, Stricker GE and Wright DG, Neutrophil-mediated endothelial injury *in vitro*. Mechanisms of cell detachment. *J Clin Invest* **68**: 1394–1403, 1981.
8. Kjellén L, Oldberg A, Rubin K and Höök M, Binding of heparin and heparan sulphate to rat liver cells. *Biochem Biophys Res Commun* **74**: 126–133, 1977.
9. Connelly JC, Skidgel RA, Schulz WW, Johnson AR and Erdös EG, Neutral endopeptidase 24.11 in human neutrophils: Cleavage of chemotactic peptide. *Proc Natl Acad Sci USA* **82**: 8737–8741, 1985.
10. Peters TJ, Muller M and DeDuve C, Lysosomes of the arterial wall. I Isolation and subcellular fractionation of cells from normal rabbit aorta. *J Exp Med* **136**: 1117–1134, 1972.
11. Schwartz D, *Méthodes Statistiques à l'Usage des Médecins et des Biologistes*. Flammarion Médecine Sciences, Paris, 1969.
12. Brestel EP and McClain EJ, A mechanism for inhibition of luminol-dependent neutrophil chemiluminescence by polyanions. *J Immunol* **131**: 2515–2519, 1983.
13. Wilhem DL, Inflammation and healing. In: *Pathology* (Eds. Anderson WAD and Kissane JM), pp. 25–89. CV Mosby Company, Saint-Louis, 1977.

Metabolism of diethylnitrosamine by microsomes of human respiratory nasal mucosa and liver

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Although the incidence of nasal tumors in the human population is low, certain individuals such as the workers in the leather or wood industry experience a higher risk of nasal cancer [1].

Nitrosamines can cause tumors in different tissues including the nasal cavity of experimental rodents [2–4]. They may also have a role in inducing cancer in the human nose as well as other respiratory tracts. The volatile diethylnitrosamine (DEN) present in air, water, foods and tobacco smoke [5] can be inhaled or absorbed from other tissues and readily passed to the nose. In rodents DEN, in addition

to other nitrosamines administered i.p., reaches the nose in a few minutes [4] and is metabolically activated via the cytochrome P-450 dependent monooxygenases system [4, 6, 7] in the nasal mucosa.

We have recently described the presence of drug-metabolizing enzymes in human respiratory nasal mucosa [8]. In the present study we have investigated whether DEN can be deethylated by human nasal mucosa microsomal enzymes. The results have been compared with those obtained with microsomal preparations from human liver.

Methods

Nasal microsomes were prepared as previously described [6, 8] from nasal respiratory membranes obtained after surgical removal for therapeutic purposes from male and female humans affected by hypertrophy of the inferior turbinates. Nasal tissues were removed from patients and frozen in liquid N₂ within 10–15 min after removal and stored at –80° until assayed.

Washed microsomes were resuspended in 20% glycerol/Tris buffer (50 mM, pH 7.4), 1 mM EDTA. Wedge biopsies of livers were obtained from patients undergoing colecystectomy. The surplus of the material required for histological analysis was made available for our studies. Tissue (about 0.5 g) was promptly frozen and stored at –80°.

Liver and mucosa microsomal fractions were prepared as previously described for rat liver [6, 9]. The extent of oxidative deethylation of DEN was determined by measuring acetaldehyde formation by HPLC according to Farrelly [10] as previously reported in detail [6]. Microsomal proteins were assayed by the method of Lowry *et al.* [11].

Results and discussion

Previous studies have shown that the major metabolite of DEN is acetaldehyde in both rodent [6, 7, 12] and human [13] liver. Acetaldehyde is formed by oxidation of DEN at the methylene carbon atom to form the α -hydroxynitrosamine. This compound has a very short lifetime under physiological conditions and it rearranges to ethyldiazohydroxide which can react with cellular constituents or with water to form ethyl alcohol or ethylene [7].

The results summarized in Table 1 show the presence of DEN-deethylase (DENd) activity in microsomes prepared from human nasal mucosa. No acetaldehyde formation was observed either when nasal mucosa microsomes previously boiled for 5 min in a boiling water bath were used in the complete incubation system or when NADPH or its regenerating system was omitted from the incubation mixture.

Table 2 shows the oxidative dealkylation of DEN by liver monooxygenases using two substrate concentrations.

In human liver DENd activities obtained at a concentration of either 1 mM or 50 mM DEN, are similar or

higher than those previously reported [13] although our data show a stronger dependence of the rates on the DEN concentration. These data suggest that besides a human P-450j isozyme (inducible by ethanol) reported to work specifically at low DEN concentration [13], DEN is also appreciably metabolized in human liver by other P-450 forms. Comparison between the DEN dealkylation rates in nasal mucosa and liver suggests that the interindividual variation of the rate of acetaldehyde formation in human liver is small, whereas that in nasal mucosa is greater. The observed variation could reflect the different ability of respiratory mucosa to metabolize DEN as has been shown for other substrates of human nasal P-450 dependent monooxygenase [8]. However at low DEN concentration the interindividual variation of DENd was smaller. The low DENd activity at a DEN concentration of 1 mM can indicate that in the respiratory section of human nose a very low amount of P-450j occur constitutively. This P-450j has been suggested to metabolize selectively DEN at low concentration in human liver [13]. By contrast, rabbit nasal mucosa contain a substantial amount of P-450j [14]. Other P-450 forms with less affinity towards DEN would be responsible for DEN metabolism when added at 50 mM. A striking result is the similarity of DENd activity in nasal mucosa and human liver.

Keeping in mind that the P-450 content is reported to be about 0.2–0.8 nmol/mg protein in human liver [15] and 0.026 nmol/mg protein in human respiratory nasal mucosa [8], the DEN-deethylation activity of nasal mucosa if expressed per nmol P-450 is 10–25 times higher than that of human liver. In rabbit [7] and hamster [6] the nasal DENd activity was 9- and 2-fold higher, respectively, than in the liver. By contrast, nasal mucosa and liver of rat showed similar DENd activities [6].

In conclusion we have found that the human respiratory nasal mucosa is able to catalyse the oxidative deethylation of DEN at a rate similar to liver. Since human nasal metabolism in the olfactory region, or in certain specific cells, is expected to be higher than in the respiratory tract by analogy with what has been found for other mammalian species [3], a role of human nasal mucosa in the bioactivation *in situ* of DEN by direct alkylation through the

Table 1. DEN-deethylase activity of human nasal mucosa microsomes^a

Patient number	Sex ^b	Age (years)	Smoking habits	Rates of CH ₃ CHO formation at DEN concentrations of	
				1 mM	50 mM
12	F	16	non-smoker	0.07	0.15
13	F	17	non-smoker	0.04	2.46
14	M	30	smoker	0.15	1.01
15	F	24	non-smoker	0.12	0.63

^a The assay mixture contained respiratory nasal mucosa microsomes (1.5 mg protein/ml) and DEN as indicated in a total volume of 2.5 ml. After 30 min of incubation at 37°, acetaldehyde formed was determined by HPLC and expressed as nmol/mg protein per min.

^b M: male; F: female.

Table 2. DEN-deethylase activity of human liver microsomes^a

Patient number	Sex ^b	Age (years)	Rates of CH ₃ CHO formation at DEN concentrations of	
			1 mM	50 mM
1	F	48	n.d.	0.8
2	M	48	1.3	4.2
3	F	57	0.5	2.1
4	F	63	1.8	4.1
5	F	62	0.6	2.3

^a The assay mixture contained liver microsomes (0.8 mg protein/ml) and DEN as indicated in a total volume of 2.0 ml. After 30 min of incubation at 37°, acetaldehyde formed was determined by HPLC and expressed as nmol/mg protein per min.

^b M: male; F: female; n.d. = not determined.

ethylating intermediates or by the formation of the nasal carcinogens acetaldehyde [16] or ethylene seems possible.

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REFERENCES

1. Buiatti M, Geddes M, Carnevale F and Merier E, Nasal cavity and paranasal sinus tumors in woodworkers and shoemakers in Italy compared to other countries. In: *Nasal Tumors in Animals and Men* (Eds. Reznik G and Stinson SF), Vol. I, pp. 111–149. CRC Press, Inc., Boca Raton, Florida, 1983.
2. Lijinsky W, Species differences in nitrosamine carcinogenesis. *J Cancer Res Clin Oncol* **108**: 46–55, 1984.
3. Dahl AR, Activation of carcinogens and other xenobiotics by nasal cytochrome P-450. In: *Microsomes and Drug Oxidation* (Eds. Boobis AR, Cold Well J, de Matteis F and Eleombe CR), pp. 299–309. Taylor and Francis, London, 1985.
4. Brittebo EB and Tjalve H, Metabolism of *N*-nitrosamines by the nasal mucosa. In: *Nasal Tumors in Animals and Men: Anatomy, Physiology and Epidemiology* (Eds. G. Reznik and S. F. Stinson), Vol. III, pp. 233–250. CRC Press Inc., Boca Raton, Florida, 1983.
5. International Agency for Research on Cancer, Some *N*-nitroso compounds. IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans. Vol. 17, pp. 83–124. IARC, Lyon, 1978.
6. Longo V, Citti L and Gervasi PG, Metabolism of diethylnitrosamine by nasal mucosa and hepatic microsomes from hamster and rat: species specificity of nasal mucosa. *Carcinogenesis* **7**: 1323–1328, 1986.
7. Ding X and Coon MJ, Cytochrome P-450-dependent formation of ethylene from *N*-nitrosoethylamine. *Drug Metab Disp* **16**: 265–269, 1988.
8. Gervasi PG, Longo V, Ursino F and Panattoni G, Drug metabolizing enzymes in respiratory mucosa of humans. Comparison with rats. In: *Proc. 6th Int. Conf. Biochemistry and Biophysics of Cytochrome P-450*. Taylor and Francis, London, in press.
9. Longo V, Citti L and Gervasi PG, Biotransformation enzymes in nasal mucosa and liver of Sprague–Dawley rats. *Toxicol Lett* **44**: 289–297, 1988.
10. Farrelly JG, A new assay for the microsomal metabolism of nitrosamines. *Cancer Res* **40**: 3241–3244, 1980.
11. Lowry OH, Rosebrough NJ, Farr AL and Randall RJ, Protein measurement with the folin phenol reagent. *J Biol Chem* **193**: 265–275, 1951.
12. Lai DY and Arcos JC, Dialkylnitrosamine bioactivation and carcinogenesis. *Life Sci* **27**: 2149–2165, 1980.
13. Yoo JH, Guengerich FP and Yung CS, Metabolism of *N*-nitrosodialkylamines by human liver microsomes. *Cancer Res* **88**: 1499–1504, 1988.
14. Ding X, Koop DR, Crump BL and Coon MJ, Immunochemical identification of cytochrome P-450 isozyme 3a (P-450 ALC) in rabbit nasal and kidney microsomes and evidence for differential induction by alcohol. *Molec Pharmacol* **30**: 370–378, 1986.
15. Kato R, Mixed function oxidases in microsomes from human liver. In: *Hepatic Cytochrome P-450 Monooxygenase System* (Eds. Schenkman JB and Kupfer D), pp. 141–145. Pergamon Press, Oxford, 1982.
16. Feron VJ, Kruysse A and Woutersen RA, Respiratory tract tumors in hamsters exposed to acetaldehyde vapor alone or simultaneously to benz(a)pyrene or diethylnitrosamine. *Eur J Cancer Clin Oncol* **18**: 13–31, 1982.

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Gonadotropin-dependent metabolism of 7,12-dimethylbenz(a)anthracene in the ovary of rhesus monkey

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7,12-Dimethylbenz(a)anthracene (DMBA)* is a potent inducer of skin and breast tumors in the rat. This compound affects several steroidogenic organs in another way. DMBA causes necrosis in the two inner zones of the rat adrenal cortex [1] and in the germinal epithelial cells in the seminiferous tubuli of the testis [2] in this same animal. The adrenal necrosis is dependent on the presence of a fully functional pituitary gland, probably related to ACTH-

induced maturity of the middle layer of the adrenal cortex and does not occur in immature animals [3].

Cell death caused by DMBA treatment may be prevented by coadministration of certain inhibitors and inducers of cytochrome P-450 [4]. Antioxidants also prevent this cytotoxicity partially, as well as preventing the extensive cytotoxic effect of 7-hydroxymethyl-12-methylbenz(a)anthracene in rat adrenal cell cultures [5], suggesting the involvement of a peroxidative mechanism, generating reactive oxygen, in the development of cellular necrosis. In addition, covalent binding of DMBA-metabolites to adrenal microsomal protein was increased 7-fold in the presence of peroxidase, indicating that, using endogenous hydrogen peroxide, peroxidases may activate

* Abbreviations used: ACTH, adrenocorticotrophic hormone; DMBA, 7,12-dimethylbenz(a)anthracene; Hepes, *N*-2-hydroxy-ethylpiperazin-*N*-2-ethane sulfonic acid; PAH, polycyclic aromatic hydrocarbons; PMSG, pregnant mare's serum gonadotropin.